

**APPLICATION INDOLE DERIVATIVE IN INFECTIOUS VIRAL DISEASES****N.K. Awasthi<sup>1</sup>****Sumit Moulekhi<sup>2</sup>**<sup>1,2</sup> Department of Chemistry B.S.N.V.PG. College Lucknow U.P. India**ABSTRACT:**

The common nitrogen-containing heterocycles, such as indoles, that have been used in the development of antiviral drugs are the main topic of our review today. The main components of pharmacological medications that have biological activity against different types of viral infections are indoles heterocycles.

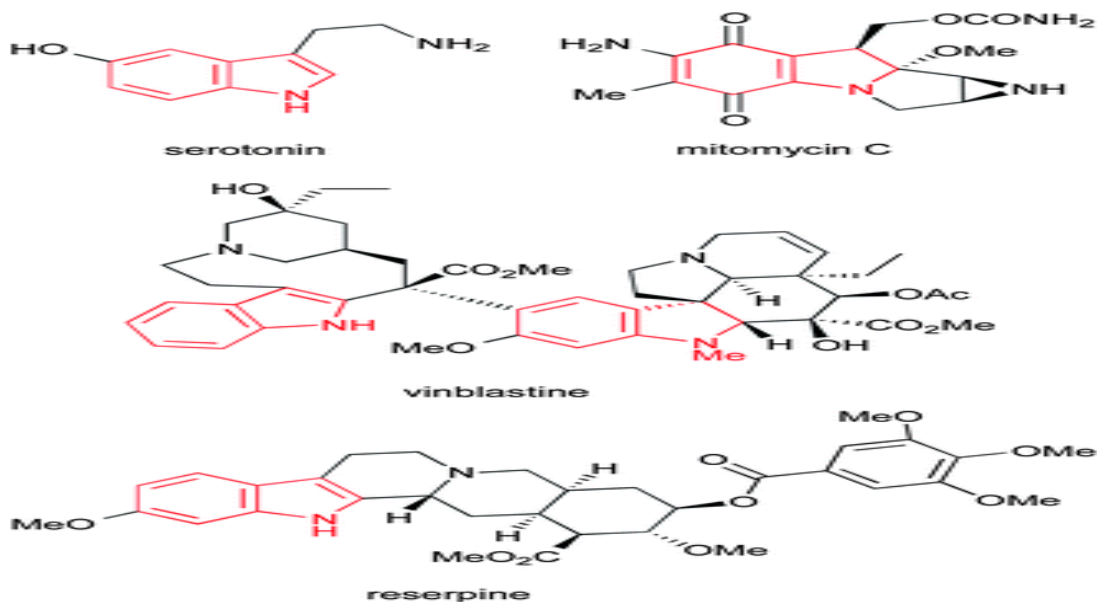
**Keywords:**

Indole, medication, antiviral design, pharmaceutical,

**INTRODUCTION:**

Infectious diseases are caused by many uncontrolled muted viruses

First of all, Millions of lives have been lost as a result of the sudden increase in infectious viral illness outbreaks in recent years. In spite of numerous established vaccinations and treatments for prevention and treatment, viruses continue to change and resurface as a threat to social cohesion, public health, and economic stability. One of the most prevalent and significant heterocycles in nature is the indole ring system. One of the most prevalent and significant heterocycles in nature is the indole ring system. found in a remarkably wide range of physiologically relevant natural substances, ranging from complex alkaloids to simple derivatives like the neurotransmitter serotonin. Indoles are ubiquitous in biological chemistry and can be found in a wide range of physiologically significant natural compounds, from simple derivatives like the neurotransmitter serotonin to complex alkaloids like the antihypertensive alkaloid reserpine and the clinically used anticancer agents' vinblastine and mitomycin C. Moreover, an indole motif is present in some significant synthetic medications, such as sumatriptan, tadalafil, rizatriptan, and fluvastatin,



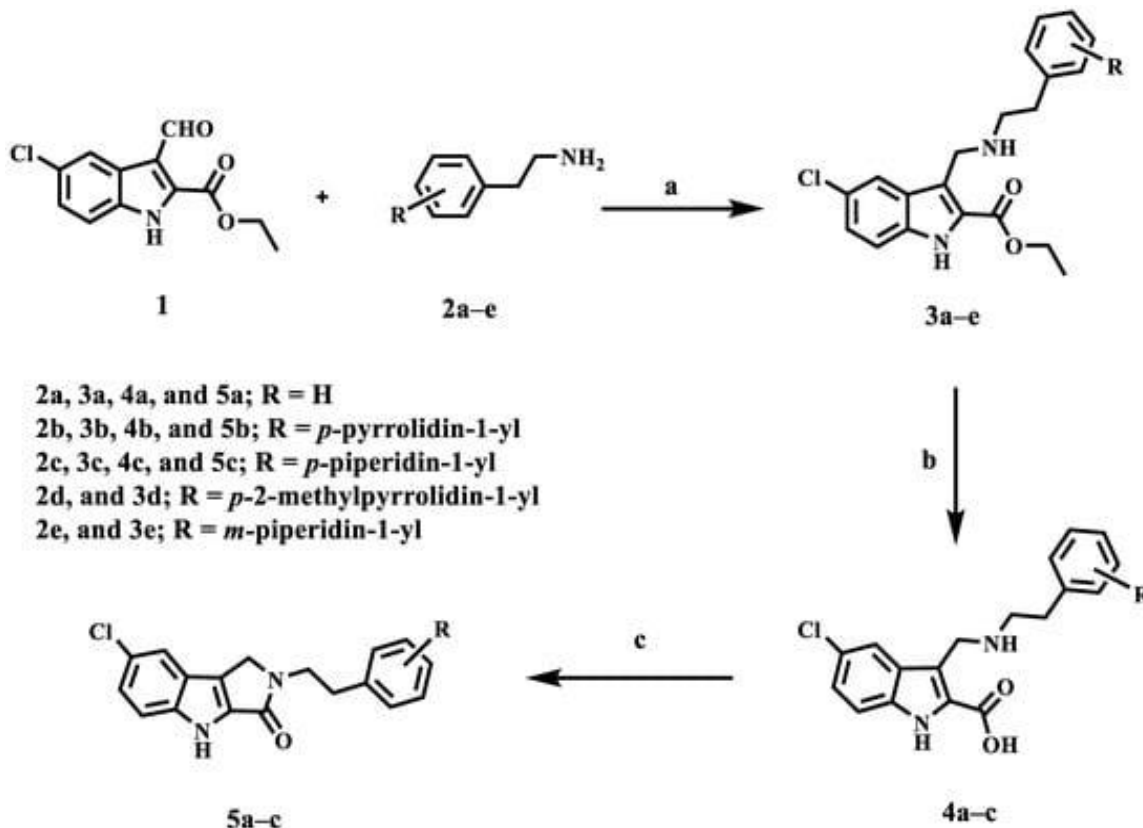
Globally, cancer has emerged as a significant public health concern due to the rising annual diagnosis rate. Unfortunately, poor selectivity, harsh side effects, and drug resistance make chemotherapy less effective when used as the main cancer treatment. Thus, it has recently been suggested that immunotherapy and newly integrated, multi-targeted medicines be used. Drug resistance, invasiveness, and cancer cell survival have all been related to kinase activity in different cell signalling pathways. Because of this, anticancer medications that target serine/threonine kinases like BRAF and kinases like the epidermal growth factor receptor (EGFR) are becoming more and more well-liked. Consequently, kinase-targeting anticancer medications are becoming more and more common. Examples of these include the epidermal growth factor receptor (EGFR) and serine/threonine kinases like BRAF. Roughly 70% of melanoma, 100% of hairy cell leukemia, and 41% of hepatocellular carcinoma have RAF mutations. In the meantime, lung, breast, and epithelial malignancies have been shown to harbour EGFR mutations including T790M and C797S, which are significant therapeutic targets. Since many types of cancer have over-activated mutant RAF/EGFR pathways, these pathways are important targets for the development of anti-cancer drugs. Conversely, the indole skeleton, which may be found in a variety of natural products and active ingredients, is one of the most well-known structures with strong anticancer activity. Many indole compounds have been found to be potent anticancer drugs thus far, and some have even been put to use in clinical settings. Several indole-based compounds with tyrosine kinase inhibitory action have been discovered in the literature research. These compounds have been shown to exhibit potential EGFR inhibitory activity and potent anticancer activity against four cancer cell lines. Additionally showing encouraging EGFR inhibitory action. It has been determined that Compound II is a dual EGFR/T790M/c-MET inhibitor that can be used to target NSCLC that is resistant. Compound II's IC<sub>50</sub> values for EGFR/T790M, EGFR/L858R, and c-MET were 0.094, 0.099, and 0.597  $\mu\text{M}$ , respectively. An EGFR TKI with a 200-fold selectivity index toward the EGFR T790M/L858R protein over wild-type EGFR is osimertinib, an indole-based medication. An EGFR TKI with a 200-fold selectivity index toward the EGFR T790M/L858R protein over wild-type EGFR is osimertinib, an indole-based medication. The FDA authorized osimertinib in 2015 for the treatment of NSCLC with EGFR T790M positivity. As EGFR-TK antiproliferative drugs, we recently reported on the

discovery of a novel class of 5-chloro-3-hydroxymethyl-indole-2-carboxamides. With an IC<sub>50</sub> value of 0.12 μM, compound (R= 4-morpholin-4-yl) was the most effective EGFR inhibitor. with an IC<sub>50</sub> value of 0.12 μM.

A series of pyrazino[1,2-*a*] indol-1(2*H*)-ones has been reported as antiproliferative agents targeting EGFR and BRAF<sup>V600E</sup>. Compound inhibits both EGFR and BRAF<sup>V600E</sup> with IC<sub>50</sub> values of 1.7 μM and 0.1 μM, respectively. Following this, a series of structural modifications to our lead compound **V** to design and synthesize a new series of pyrazino[1,2-*a*] indol-1(2*H*)-ones. Compound was the most effective derivative, with a GI<sub>50</sub> value of 1.107 μM against four cancer cell lines. inhibited EGFR with an IC<sub>50</sub> of 0.08 μM but only moderately inhibited BRAF<sup>V600E</sup> with an IC<sub>50</sub> of 0.15 μM. In another study, we describe our efforts to synthesize and optimize a novel class of potent antiproliferative agents. The antiproliferative activity of the target compounds is impressive. These compounds have a dual inhibitory effect on EGFR and BRAF<sup>V600E</sup>, with IC<sub>50</sub> values of 32 nM and 45 nM, respectively.

Motivated by the data presented above, and as part of our ongoing efforts to identify promising lead compounds for dual or multi-targeted anticancer agents, we present herein the design and synthesis of a novel class of indole-2-carboxylates, as well as 1,2-dihydropyrrolo[3,4-*b*] indol-3(4*H*)-ones, compounds, as dual EGFR/BRAF<sup>V600E</sup> inhibitors with antiproliferative activity. The new compounds will be evaluated for their safety profile by assessing their effect on the viability of human normal cell lines, while their antiproliferative activity will be evaluated against a panel of four cancer cell lines. The most potent compounds will be evaluated for their ability to inhibit wild-type EGFR (EGFR<sup>WT</sup>) and BRAF<sup>V600E</sup> as a potential mechanistic target for their antiproliferative effects. Furthermore, the most potent EGFR inhibitors will be tested for their inhibitory effect against mutant-type EGFR (EGFR<sup>T790M</sup>), and the most potent anti-BRAF agents will be tested for their anticancer effect against the LOX-IMVI melanoma cell line, which has BRAF<sup>V600E</sup> kinase overexpression. Finally, docking studies will be performed to investigate these compounds' binding interactions within the active sites of target enzymes.

The synthesis of target compounds **3a-e**, **4a-c**, and **5a-c** is depicted in **Scheme 1**. 5-chloro-3-formyl indole-2-carboxylate **1** [] was reacted with amines **2a-e** [] through reflux in ethanol followed by reduction of the intermediate imine with NaBH<sub>4</sub> under reductive-amination conditions to yield secondary amines **3a-e** which subjected to saponification with Li OH to afford a carboxylic acids **4a-c**. The structures of compounds **3a-e** and **4a-c** were confirmed using <sup>1</sup>H NMR, <sup>13</sup>C NMR, and HRESI-MS spectroscopy (Varian Inova, University of Aberdeen, Meston Building, Aberdeen AB24 3UE, UK). <sup>1</sup>H NMR spectrum of compound **3c** revealed the presence of a singlet signal δ 9.12 ppm of indole NH, the characteristic signals of ethyl group in the form of quartet at δ 4.33 ppm (2H) and triplet at δ 1.35 ppm (3H), a singlet signal at δ 4.18 ppm (2H) of CH<sub>2</sub>NH- group, and two triplets (each of 2H) at δ 2.88 and 2.74 ppm of NHCH<sub>2</sub>CH<sub>2</sub> group. Moreover, the spectrum revealed the presence of the characteristic signals of both piperidine and aromatic protons. HRESI-MS *m/z* of **3c** clad for [M + H]<sup>+</sup> C<sub>25</sub>H<sub>31</sub>ClN<sub>3</sub>O<sub>2</sub>: 440.2099, found: 440.2100. The disappearance of the characteristic signals of the ethyl group in the form quartet at δ 4.33 ppm (2H) and triplet at δ 1.35 ppm (3H) and the appearance of a singlet signal at δ 3.43 ppm (2H) corresponding to COOH and NHCH<sub>2</sub> characterize the <sup>1</sup>H NMR spectrum of **4c**.



**Scheme 1.** Synthesis of compounds **3a–e**, **4a–c**, and **5a–c**. *Reagents and conditions:* (a) NaBH<sub>4</sub>, EtOH, reflux, 12 h to rt, 1 h, 78%; (b) LiOH, THF/ H<sub>2</sub>O, 40 °C, 90%; (c) BOP, DIPEA, DMF, rt, overnight, 70%.

**Conclusion:** Using a variety of spectroscopic techniques, a new series of 5-chloro-indole-2-carboxylate and pyrrole [3,4-*b*] indol-3-one was produced and structurally described. The novel compounds exhibited strong antiproliferative properties against four human cancer cell lines, however they showed no cytotoxic effects on human normal cell lines. It was discovered that a few of the substances examined were dual inhibitors of BRAFV600E and EGFR, both of the wild type and mutant types. Using vemurafenib as a reference, molecular docking was used to look into the binding manner of the most potent antiproliferative drugs (**3a–e**) within the BRAFV600E binding site. The results showed that compound **3e** fit more snugly within the active site than the other derivatives with para-amine substituents because it had an *m*-piperidinyl substitution at the phenethyl amine moiety.

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the EGFRT790M and BRAFV600E pathways; however, additional in vitro and in vivo testing is required. Using vemurafenib as a reference, molecular docking was used to look into the binding manner of the most potent antiproliferative drugs (3a–e) within the BRAFV600E binding site. The findings demonstrated that compound 3e, which substitutes m-piperidinyI at the phenethyl amine moiety, suited the data more closely. Additionally, compounds 3b and 3e's docking experiments against EGFRT790M indicate that the ligand indole-2-carboxylate scaffold binds well, generating a combination of hydrophobic and H-bond interactions at the active site's hydrophobic pocket. Compounds 3a–e show strong pharmacokinetic and ADME properties, according to in silico ADME and pharmacokinetic prediction. After structural alterations, compounds 3b and 3e may function as anticancer drugs targeting the EGFRT790M and BRAFV600E pathways; however, additional in vitro and in vivo testing is required.

#### REFERENCES:

1. Ghosh A.K., Osswald H.L., Prato G. Recent Progress in the Development of HIV-1 Protease Inhibitors for the Treatment of HIV/AIDS. *J. Med. Chem.* 2016; 59:5172–5208. Doi: 10.1021/acs.jmedchem.5b01697. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
2. Krammer F., Palese P. Advances in the development of influenza virus vaccines. *Nat. Rev. Drug Discov.* 2015; 14:167–182. doi: 10.1038/nrd4529. [PubMed] [CrossRef] [Google Scholar]
3. Bolles M., Donaldson E., Baric R. SARS-CoV and emergent coronaviruses: Viral determinants of interspecies transmission. *Curr. Opin. Virol.* 2011;1:624–634. Doi: 10.1016/j.coviro.2011.10.012. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
4. Wang X., Zou P., Wu F., Lu L., Jiang S. Development of small-molecule viral inhibitors targeting various stages of the life cycle of emerging and re-emerging viruses. *Front. Med.* 2017; 11:449–461. Doi: 10.1007/s11684-017-0589-5. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
5. Korkmaz A., Bursal E. An in vitro and in silico study on the synthesis and characterization of novel bis(sulfonate) derivatives as tyrosinase and pancreatic lipase inhibitors. *J. Mol. Struct.* 2022;1259:132734. doi: 10.1016/j.molstruc.2022.132734. [CrossRef] [Google Scholar]
6. Cetin A., Bursal E., Türkan F. 2-methylindole analogs as cholinesterases and glutathione S-transferase inhibitors: Synthesis, biological evaluation, molecular docking, and pharmacokinetic studies. *Arab. J. Chem.* 2021;14:103449. doi: 10.1016/j.arabjc.2021.103449. [CrossRef] [Google Scholar]
7. Zhang M.Z., Chen Q., Yang G.F. A review on recent developments of indole-containing antiviral agents. *Eur. J. Med. Chem.* 2015;89:421–441. doi: 10.1016/j.ejmech.2014.10.065. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
8. Mohana Roopan S., Sompalle R. Synthetic chemistry of pyrimidines and fused pyrimidines: A review. *Synth. Commun.* 2016;46:645–672. doi: 10.1080/00397911.2016.1165254. [CrossRef] [Google Scholar]
9. Fascio M.L., Errea M.I., D'Accorso N.B. Imidazothiazole and related heterocyclic systems. Synthesis, chemical and biological properties. *Eur. J. Med. Chem.* 2015;90:666–683. doi: 10.1016/j.ejmech.2014.12.012. [PubMed] [CrossRef] [Google Scholar]
10. Shalini K., Sharma P.K., Kumar N. Imidazole and its biological activities: A review. *Der Chem. Sin.* 2010;1:36–47. [Google Scholar]
11. Singh N., Pandurangan A., Rana K., Anand P., Ahamad A., Tiwari A.K. Benzimidazole: A short review of their antimicrobial activities. *Int. Curr. Pharm. J.* 2012;1:110–118. doi: 10.3329/icpj.v1i5.10284. [CrossRef] [Google Scholar]
12. Gupta V., Kant V. A review on biological activity of imidazole and thiazole moieties and their derivatives. *Sci. Int.* 2013;1:253–260. doi: 10.17311/sciintl.2013.253.260. [CrossRef] [Google Scholar]



13. Bhardwaj V., Gumber D., Abbot V., Dhiman S., Sharma P. Pyrrole: A resourceful small molecule in key medicinal hetero-aromatics. *RSC Adv.* 2015;5:15233–15266. doi: 10.1039/C4RA15710A. [[CrossRef](#)] [[Google Scholar](#)]
14. Sharma V., Chitranshi N., Agarwal A.K. Significance and biological importance of pyrimidine in the microbial world. *Int. J. Med. Chem.* 2014;2014:1–31. doi: 10.1155/2014/202784. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
15. Raffa D., Maggio B., Raimondi M.V., Cascioferro S., Plescia F., Cancemi G., Daidone G. Recent advanced in bioactive systems containing pyrazole fused with a five membered heterocycle. *Eur. J. Med. Chem.* 2015;97:732–746. doi: 10.1016/j.ejmech.2014.12.023. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
16. Marella A., Tanwar O.P., Saha R., Ali M.R., Srivastava S., Akhter M., Shaquiquzzaman M., Alam M.M. Quinoline: A versatile heterocyclic. *Saudi Pharm. J.* 2013;21:1–12. doi: 10.1016/j.jsps.2012.03.002. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
17. Sundberg R.J. *Indoles*. Academic Press; London, UK: San Diego, CA, USA: 1996. [[Google Scholar](#)]
18. Hassam M., Basson A.E., Liotta D.C., Morris L., van Otterlo W.A.L., Pelly S.C. Novel Cyclopropyl-Indole Derivatives as HIV Non-Nucleoside Reverse Transcriptase Inhibitors. *ACS Med. Chem. Lett.* 2012;3:470–475. doi: 10.1021/ml3000462. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
19. Lavanchy D. Evolving epidemiology of hepatitis C virus. *Clin. Microbiol. Infect.* 2011;17:107–115. doi: 10.1111/j.1469-0691.2010.03432.x. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
20. Han Z., Liang X., Wang Y., Qing J., Cao L., Shang L., Yin Z. The discovery of indole derivatives as novel hepatitis C virus inhibitors. *Eur. J. Med. Chem.* 2016;116:147–155. doi: 10.1016/j.ejmech.2016.03.062. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
21. Chen J.J., Gershon A.A., Li Z., Cowles R.A., Gershon M.D. Varicella zoster virus (VZV) infects and establishes latency in enteric neurons. *J. Neurovirol.* 2011;17:578–589. doi: 10.1007/s13365-011-0070-1. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
22. Musella S., di Sarno V., Ciaglia T., Sala M., Spensiero A., Scala M.C., Ostacolo C., Andrei G., Balzarini J., Snoeck R., et al. Identification of an indol-based derivative as potent and selective varicella zoster virus (VZV) inhibitor. *Eur. J. Med. Chem.* 2016;124:773–781. doi: 10.1016/j.ejmech.2016.09.014. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
23. Topalis D., Gillemot S., Snoeck R., Andrei G. Thymidine kinase and protein kinase in drug-resistant herpesviruses: Heads of a Lernaean Hydra. *Drug Resist. Updat.* 2018;37:1–16. doi: 10.1016/j.drug.2018.01.003. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
24. Thanigaimalai P., Konno S., Yamamoto T., Koiwai Y., Taguchi A., Takayama K., Yakushiji F., Akaji K., Chen S.E., Naser-Tavakolian A., et al. Development of potent dipeptide-type SARS-CoV 3CL protease inhibitors with novel P3 scaffolds: Design, synthesis, biological evaluation, and docking studies. *Eur. J. Med. Chem.* 2013;68:372–384. doi: 10.1016/j.ejmech.2013.07.037. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
25. Khan R.J., Jha R.K., Amera G.M., Jain M., Singh E., Pathak A., Singh R.P., Muthukumaran J., Singh A.K. Targeting SARS-CoV-2: A systematic drug repurposing approach to identify promising inhibitors against 3C-like proteinase and 2'-O-ribose methyltransferase. *J. Biomol. Struct. Dyn.* 2021;39:2679–2692. doi: 10.1080/07391102.2020.1753577. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
26. Thanigaimalai P., Konno S., Yamamoto T., Koiwai Y., Taguchi A., Takayama K., Yakushiji F., Akaji K., Kiso Y., Kawasaki Y., et al. Design, synthesis, and biological evaluation of novel dipeptide-type SARS-CoV 3CL protease inhibitors: Structure-activity relationship study. *Eur. J. Med. Chem.* 2013;65:436–447. doi: 10.1016/j.ejmech.2013.05.005. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

27. Dai W., Zhang B., Jiang X.-M., Su H., Li J., Zhao Y., Xie X., Jin Z., Peng J., Liu F., et al. Structure-based design of antiviral drug candidates targeting the SARS-CoV-2 main protease. *Science*. 2020;368:1331–1335. doi: 10.1126/science.abb4489. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
28. Jones R., Bean G. *The Chemistry of Pyrroles*. Academic Press; London, UK: 1997. [Google Scholar]
29. Curreli F., Kwon Y.D., Belov D.S., Ramesh R.R., Kurkin A.V., Altieri A., Kwong P.D., Debnath A.K. Synthesis, Antiviral Potency, in Vitro ADMET, and X-ray Structure of Potent CD4 Mimics as Entry Inhibitors That Target the Phe43 Cavity of HIV-1 gp120. *J. Med. Chem.* 2017;60:3124–3153. doi: 10.1021/acs.jmedchem.7b00179. [PubMed] [CrossRef] [Google Scholar]
30. Checkley M.A., Luttge B.G., Freed E.O. HIV-1 envelope glycoprotein biosynthesis, trafficking, and incorporation. *J. Mol. Biol.* 2011;410:582–608. doi: 10.1016/j.jmb.2011.04.042. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
31. Ezquerra J., Pedregal C., Rubio A., Valenciano J., Navio J.L.G., Alvarez-Builla J., Vaquero J.J. General method for the synthesis of 5-arylpyrrole-2-carboxylic acids. *Tetrahedron Lett.* 1993;34:6317–6320. doi: 10.1016/S0040-4039(00)73741-5. [CrossRef] [Google Scholar]
32. Kinchington P.R., Leger A.J., Guedon J.M., Hendricks R.L. Herpes simplex virus and varicella zoster virus, the house guests who never leave. *Herpesviridae*. 2012;3:5. doi: 10.1186/2042-4280-3-5. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
33. Hilmy K.M., Soliman D.H., Shahin E.B., Abd Alhameed R. Synthesis and molecular modeling study of novel pyrrole Schiff Bases as anti-HSV-1 agents. *Life Sci. J.* 2012;9:736–745. [Google Scholar]
34. Lin M.I., Su B.H., Lee C.H., Wang S.T., Wu W.C., Dangate P., Wang S.Y., Huang W.I., Cheng T.J., Lin O.A., et al. Synthesis and inhibitory effects of novel pyrimido-pyrrolo-quinoxalinedione analogues targeting nucleoproteins of influenza A virus H1N1. *Eur. J. Med. Chem.* 2015;102:477–486. doi: 10.1016/j.ejmech.2015.08.016. [PubMed] [CrossRef] [Google Scholar]
35. Turrell L., Lyall J.W., Tiley L.S., Fodor E., Vreede F.T. The role and assembly mechanism of nucleoprotein in influenza A virus ribonucleoprotein complexes. *Nat. Commun.* 2013;4:1591. doi: 10.1038/ncomms2589. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
36. Brown D.J. *The Pyrimidines*. John Wiley & Sons; New York, NY, USA: 1994. pp. 1–8. [Google Scholar]
37. Dinakaran V.S., Bomma B., Srinivasan K.K. Fused pyrimidines: The heterocycle of diverse biological and pharmacological significance. *Der Pharma Chem.* 2012;4:255–265. [Google Scholar]
38. Malancona S., Mori M., Fezzardi P., Santoriello M., Basta A., Nibbio M., Kovalenko L., Speziale R., Battista M.R., Cellucci A., et al. 5,6-Dihydroxypyrimidine Scaffold to Target HIV-1 Nucleocapsid Protein. *ACS Med. Chem. Lett.* 2020;11:766–772. doi: 10.1021/acsmchemlett.9b00608. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
39. Levin J.G., Mitra M., Mascarenhas A., Musier-Forsyth K. Role of HIV-1 nucleocapsid protein in HIV-1 reverse transcription. *RNA Biol.* 2010;7:754–774. doi: 10.4161/rna.7.6.14115. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
40. Mohamed M.S., Sayed A.I., Khedr M.A., Soror S.H. Design, synthesis, assessment, and molecular docking of novel pyrrolopyrimidine (7-deazapurine) derivatives as non-nucleoside hepatitis C virus NS5B polymerase inhibitors. *Bioorg. Med. Chem.* 2016;24:2146–2157. doi: 10.1016/j.bmc.2016.03.046. [PubMed] [CrossRef] [Google Scholar]
41. Zhao C., Wang Y., Ma S. Recent advances on the synthesis of hepatitis C virus NS5B RNA-dependent RNA-polymerase inhibitors. *Eur. J. Med. Chem.* 2015;102:188–214. doi: 10.1016/j.ejmech.2015.07.046. [PubMed] [CrossRef] [Google Scholar]

42. Mohamed S.F., Flefel E.M., Amr Ael G., Abd El-Shafy D.N. Anti-HSV-1 activity and mechanism of action of some new synthesized substituted pyrimidine, thiopyrimidine and thiazolopyrimidine derivatives. *Eur. J. Med. Chem.* 2010;45:1494–1501. doi: 10.1016/j.ejmech.2009.12.057. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
43. Wang M., Cao R., Zhang L., Yang X., Liu J., Xu M., Shi Z., Hu Z., Zhong W., Xiao G. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res.* 2020;30:269–271. doi: 10.1038/s41422-020-0282-0. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
44. Zhang L., Zhou R. Structural Basis of the Potential Binding Mechanism of Remdesivir to SARS-CoV-2 RNA-Dependent RNA Polymerase. *J. Phys. Chem. B.* 2020;124:6955–6962. doi: 10.1021/acs.jpcc.0c04198. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
45. Grein J., Ohmagari N., Shin D., Diaz G., Asperges E., Castagna A., Feldt T., Green G., Green M.L., Lescure F.X., et al. Compassionate Use of Remdesivir for Patients with Severe COVID-19. *N. Engl. J. Med.* 2020;382:2327–2336. doi: 10.1056/NEJMoa2007016. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
46. Ansari A., Ali A., Asif M., Shamsuzzaman Review: Biologically active pyrazole derivatives. *New J. Chem.* 2017;41:16–41. doi: 10.1039/C6NJ03181A. [[CrossRef](#)] [[Google Scholar](#)]
47. Eicher T., Hauptmann S., Speicher A. *The Chemistry of Heterocycles: Structure, Reactions, Syntheses, and Applications.* Wiley-VCH; Weinheim, Germany: 2003. [[Google Scholar](#)]
48. Christen M.T., Menon L., Myshakina N.S., Ahn J., Parniak M.A., Ishima R. Structural basis of the allosteric inhibitor interaction on the HIV-1 reverse transcriptase RNase H domain. *Chem. Biol. Drug Des.* 2012;80:706–716. doi: 10.1111/cbdd.12010. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
49. Messori A., Corona A., Madia V.N., Saccoliti F., Tudino V., De Leo A., Scipione L., De Vita D., Amendola G., Di Maro S., et al. Pyrrolyl Pyrazoles as Non-Diketo Acid Inhibitors of the HIV-1 Ribonuclease H Function of Reverse Transcriptase. *ACS Med. Chem. Lett.* 2020;11:798–805. doi: 10.1021/acsmchemlett.9b00617. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
50. Blyth C.C., Jacoby P., Effler P.V., Kelly H., Smith D.W., Robins C., Willis G.A., Levy A., Keil A.D., Richmond P.C. Effectiveness of trivalent flu vaccine in healthy young children. *Pediatrics.* 2014;133:e1218–e1225. doi: 10.1542/peds.2013-3707. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
51. David J.K., Amber F. Extremely low vaccine effectiveness against influenza H3N2 in the elderly during the 2012/2013 flu season. *J. Infect. Dev. Ctries.* 2013;7:299–301. doi: 10.3855/jidc.3544. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
52. Meng F.J., Sun T., Dong W.Z., Li M.H., Tuo Z.Z. Discovery of Novel Pyrazole Derivatives as Potent Neuraminidase Inhibitors against Influenza H1N1 Virus. *Arch. Pharm.* 2016;349:168–174. doi: 10.1002/ardp.201500342. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
53. Jia H., Bai F., Liu N., Liang X., Zhan P., Ma C., Jiang X., Liu X. Design, synthesis and evaluation of pyrazole derivatives as non-nucleoside hepatitis B virus inhibitors. *Eur. J. Med. Chem.* 2016;123:202–210. doi: 10.1016/j.ejmech.2016.07.048. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
54. Prajapati S.M., Patel K.D., Vekariya R.H., Panchal S.N., Patel H.D. Recent advances in the synthesis of quinolines: A review. *RSC Adv.* 2014;4:24463–24476. doi: 10.1039/C4RA01814A. [[CrossRef](#)] [[Google Scholar](#)]
55. Overacker R.D., Banerjee S., Neuhaus G.F., Milicevic Sephton S., Herrmann A., Strother J.A., Brack-Werner R., Blakemore P.R., Loesgen S. Biological evaluation of molecules of the azaBINOL class as



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antiviral agents: Inhibition of HIV-1 RNase H activity by 7-isopropoxy-8-(naphth-1-yl)quinoline. *Bioorg. Med. Chem.* 2019;27:3595–3604. doi: 10.1016/j.bmc.2019.06.044. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

56. Shah U., Jayne C., Chackalamannil S., Velázquez F., Guo Z., Buevich A., Howe J.A., Chase R., Soriano A., Agrawal S., et al. Novel Quinoline-Based P2–P4 Macrocyclic Derivatives As Pan-Genotypic HCV NS3/4a Protease Inhibitors. *ACS Med. Chem. Lett.* 2014;5:264–269. doi: 10.1021/ml400466p. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

57. Schiering N., D'Arcy A., Villard F., Simic O., Kamke M., Monnet G., Hassiepen U., Svergun D.I., Pulfer R., Eder J., et al. A macrocyclic HCV NS3/4A protease inhibitor interacts with protease and helicase residues in the complex with its full-length target. *Proc. Natl. Acad. Sci. USA.* 2011;108:21052–21056. doi: 10.1073/pnas.1110534108. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

58. Wang W., Yin R., Zhang M., Yu R., Hao C., Zhang L., Jiang T. Boronic Acid Modifications Enhance the Anti-Influenza A Virus Activities of Novel Quindoline Derivatives. *J. Med. Chem.* 2017;60:2840–2852. doi: 10.1021/acs.jmedchem.6b00326. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

59. Liu J., Cao R., Xu M., Wang X., Zhang H., Hu H., Li Y., Hu Z., Zhong W., Wang M. Hydroxychloroquine, a less toxic derivative of chloroquine, is effective in inhibiting SARS-CoV-2 infection in vitro. *Cell Discov.* 2020;6:16. doi: 10.1038/s41421-020-0156-0. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

60. Pereira B.B. Challenges and cares to promote rational use of chloroquine and hydroxychloroquine in the management of coronavirus disease 2019 (COVID-19) pandemic: A timely review. *J. Toxicol. Environ. Health B Crit. Rev.* 2020;23:177–181. doi: 10.1080/10937404.2020.1752340. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]